

(19)



Europäisches Patentamt  
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(11)

EP 1 172 378 A1

(12)

## EUROPEAN PATENT APPLICATION

(43) Date of publication:  
16.01.2002 Bulletin 2002/03

(51) Int Cl.7: C07K 16/18, A61K 39/395,  
C07K 1/22, G01N 33/68,  
A61P 25/28

(21) Application number: 01114825.1

(22) Date of filing: 28.06.2001

(84) Designated Contracting States:  
AT BE CH CY DE DK ES FI FR GB GR IE IT LI LU  
MC NL PT SE TR

Designated Extension States:  
AL LT LV MK RO SI

(30) Priority: 12.07.2000 EP 00115141

(71) Applicants:

- Dodel, Richard, Dr.  
35039 Marburg (DE)

• Du, Yansheng, Dr.  
Carmel, Indiana 46033 (US)

(72) Inventors:

- Dodel, Richard, Dr.  
35039 Marburg (DE)
- Du, Yansheng, Dr.  
Carmel, Indiana 46033 (US)

(74) Representative: Schwabe - Sandmair - Marx  
Stuntzstrasse 16  
81677 München (DE)

### (54) Human beta-amyloid antibody and use thereof for treatment of alzheimer's disease

(57) The present invention provides according to the first aspect thereof a human anti- $\beta$ -amyloid antibody obtained by purification from a human IgG-containing bodyfluid by  $\text{A}\beta$ -affinity chromatography. In a second aspect the invention provides a method of purification of an anti- $\text{A}\beta$ -amyloid antibody, said method comprising the steps of obtaining a human IgG-containing bodyfluid, subject-

ing the bodyfluid obtained to an  $\text{A}\beta$ -affinity chromatography, and recovering the purified anti- $\text{A}\beta$  antibody from the chromatography medium. Finally the invention provides for use of the above anti- $\text{A}\beta$  antibody for diagnosing and/or treating amyloid associated diseases, especially Alzheimer's disease and for a pharmaceutical composition comprising said antibody for treatment of Alzheimer's disease.

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## Description

[0001] The present invention relates to a human  $\beta$ -amyloid antibody, a method of purification thereof and the use of this  $\beta$ -amyloid antibody in treatment of amyloid associated diseases, especially Alzheimer's Disease.

## BACKGROUND

[0002] Alzheimer's disease is a progradient disease initially manifesting itself with partial amnesia, and later restlessness, dysorientation, aphasia, agnosia or apraxia (cognitive decline), dementia and sometimes euphoria or depressions. The disease typically starts at 40 to 90 years of age and predominantly affects females. As to its occurrence, estimations are about 5 % of the population above 65 years age. Alzheimer thus constitutes a major problem in industrialised countries.

[0003] In Alzheimer's disease brain region-specific amyloid deposition is a key neuropathological feature which is accompanied by astrogliosis, microgliosis, cytoskeletal changes, and synaptic loss. These pathological alterations are thought to be linked to the cognitive decline and dementia which defines the disease. These neuritic depositions or plaques and neurofibrillary tangles comprise the major neuropathological changes associated with Alzheimer's disease. Although other neuropathological changes have been linked to Alzheimer's disease, evidence indicates that they are as well somehow related to the classical lesions.

[0004] Neuritic plaques are spherical, multicellular lesions that are usually found in moderate or large numbers in limbic structures and association neocortex. The plaques contain extracellular deposits of  $\beta$ -amyloid protein ( $A\beta$ ) that include abundant amyloid fibrils intermixed with non-fibrillar forms of this peptide. The major protein constituent of plaques is the  $\beta$ -amyloid protein ( $A\beta$ ). Neuritic plaques have degenerating axons and dendrites within and intimately surrounding the plaque. Such plaques also contain variable numbers of activated microglia that are often situated within and near the fibrillar amyloid core, as well as reactive astrocytes surrounding the core.

[0005] The major constituent of the plaque, the  $\beta$ -amyloid protein, arises from a larger precursor protein, the amyloid precursor protein (APP). The amyloid precursor protein (APP) refers to a group of ubiquitously expressed proteins whose heterogeneity arises from both alternative splicing and posttranslational processes. Cleavage of APP in its COOH-terminal region in the transmembrane domain by  $\beta$ -secretase and  $\gamma$ -secretase results in the formation of the  $\beta$ -amyloid protein.

[0006]  $A\beta$  is secreted continuously by normal cells and can be detected as a circulating peptide in the plasma and cerebrospinal fluid (CSF) of healthy humans. In Alzheimer's disease it is thought that increased production of  $A\beta$  and/or a decreased metabolism of  $A\beta$  may

lead to plaque deposition and consecutively to the neuropathological changes associated with Alzheimer's disease. Evidence for the role of  $A\beta$  in Alzheimer's disease include the observation that missense mutations in the APP have been found to be the cause of familial Alzheimer disease cases.

[0007] Several endogenous substrates, including apolipoprotein E have been shown to be associated with plaque formation. In transgenic mice APPV717F (PDAPP) the lack of the apolipoprotein E gene (apoE-knock-out mice) results in the absence of amyloid plaque deposition (Games et al., Nature 1995; Bales et al., Nat Genet 1997). These transgenic mice (PDAPP) normally develop amyloid plaques in an age-dependent manner starting at three months of age.

[0008] Schenk and coworkers (Schenk et al, Nature 400:173, 1999) investigated the plaque burden in the PDAPP-mice following an immunization treatment. PDAPP-mice were immunised with pre-aggregated  $A\beta$  for different time periods using Freud's adjuvants. Plaque deposition in these mice decreased significantly following the immunization treatment. Sham-mice did not show a decrease in plaque deposition.

[0009] Treatment of APPV717F transgenic mice with antibodies raised against  $A\beta$  was also reported to attenuate amyloid plaque formation, neuritic dystrophy and astrogliosis in younger mice as well as to decrease plaque burden in older mice. However, the finding could not be verified in other mice.

[0010] Despite of the above knowledge no therapy for amyloid associated diseases, especially Alzheimer's disease is available up to today. However, an effective therapy for Alzheimer's disease would be highly desirable because of its broad spread occurrence.

[0011] It is therefore an object of the present invention to provide such therapy of and/or means for diagnosing amyloid associated diseases, especially Alzheimer's disease.

## 40 SUMMARY OF THE INVENTION

[0012] The above object can be solved by a human anti- $\beta$ -amyloid antibody and a pharmaceutical composition comprising the same as stipulated in the appending claims.

[0013] More in detail the present invention according to the first aspect thus provides a human anti- $\beta$ -amyloid antibody obtained by purification from a human IgG-containing bodyfluid by  $A\beta$ -affinity chromatography.

[0014] In a second aspect the invention provides a method of purification of an anti- $A\beta$ -amyloid antibody, said method comprising the steps of obtaining a human IgG-containing bodyfluid, subjecting the bodyfluid obtained to an  $A\beta$ -affinity chromatography, and recovering the purified anti- $A\beta$  antibody from the chromatography medium.

[0015] Finally the invention provides for use of the above anti- $A\beta$  antibody for diagnosing (with a special

developed ELISA) and/or treating amyloid associated diseases, especially Alzheimer's disease and for a pharmaceutical composition comprising said antibody for treatment of amyloid associated diseases, especially Alzheimer disease and manufacture thereof.

#### DETAILED DESCRIPTION OF THE INVENTION

[0016] The applicants have now found that naturally occurring A $\beta$  antibodies exist in biologically relevant fluids i.e. CSF and plasma, and that levels of these antibodies differ between normal age-matched healthy controls and AD patients. Based on these findings it was concluded and then supported by experiments that the antibody can be used for diagnosis and treatment of amyloid associated diseases and especially of Alzheimer's disease. In the context of this specification the terms "anti-A $\beta$  antibodies" and "A $\beta$  antibodies" are used interchangeably to designate the antibody of the invention.

[0017] In lumbar CSF samples which included 49 age-matched non-demented individuals with no family history of cognitive impairment and 60 individuals with confirmed AD, detection of CSF A $\beta$  antibody levels was determined utilising an ELISA assay in which the A $\beta$  peptide was used as the capture ligand (see below).

[0018] Human anti-A $\beta$  antibody was detected in CSF samples in both of the populations studied. It was confirmed that the A $\beta$  antibody activity detected by the ELISA represents antibodies specific to A $\beta$  by absorbing the activity with protein-A Agarose, and A $\beta_{1-40}$  or A $\beta_{1-42}$ . However, no interaction was found between this A $\beta$ -antibody and A $\beta_{40-1}$  or unrelated neuropeptides such as neuropeptide F or neuropeptide Y. The mean level of A $\beta$  antibody in the Alzheimer's disease group was 30% lower than controls (control: 370±39, AD: 276±27; p < 0.05, one way ANOVA).

[0019] These data demonstrate that an antibody directed against A $\beta$  (anti-A $\beta$  antibody or short: A $\beta$  antibody) is present in physiologically relevant concentrations in human fluids, like CSF and serum. Antibody titres are significantly higher in control subjects than AD patients. The generation of naturally occurring A $\beta$ -antibodies and subsequent A $\beta$  / antibody complex formation, may be involved in the normal clearance of A $\beta$  peptide(s), which serves to reduce A $\beta$  deposition and neuritic plaque formation.

[0020] The lower titres of A $\beta$  antibody found in more than 50% of the AD patients investigated in this study compared to controls suggest that reduced A $\beta$  antibody generation and/or complex formation contributes to an abnormal (i.e. reduced) clearance function. Similar clearance problems may occur in other neurodegenerative diseases or amyloid associated diseases such as primary and secondary amyloidoses. The present invention thus pertains to treatment and diagnosis of these other amyloid associated diseases as well.

[0021] Based on the above hypothesis the treatment with antibodies against A $\beta$  i.e. A $\beta$  antibodies is a new

strategy to treat diseases associated with amyloid deposition. These treatments include the increase of A $\beta$ -antibody levels by using immunoglobulins (IgG), preferably human IgG with high titres of A $\beta$  antibodies or using anti-A $\beta$  antibodies purified from human IgG containing fluids. The present invention also encompasses use of antibody fragments (Fab etc.) as long as complex formation can be achieved.

[0022] Thus, the present invention relates to a human anti- $\beta$ -amyloid antibody (A $\beta$ -antibody) obtained by purification from a human IgG-containing bodyfluid by A $\beta$ -affinity chromatography. Preferably the human anti-A $\beta$  antibody belongs to the class of immunoglobulines G (IgG) and does not recognise A $\beta_{40-1}$ , neuropeptide F, neuropeptide Y, and Amylin, and specifically recognises one or more of A $\beta_{1-40}$ , A $\beta_{1-42}$ , and A $\beta_{25-35}$ , and preferably recognises all of A $\beta_{1-40}$ , A $\beta_{1-42}$ , and A $\beta_{25-35}$ .

[0023] According to a second embodiment the present invention relates to a method of purification of an anti-A $\beta$ -antibody comprising the steps of obtaining a human IgG-containing bodyfluid, subjecting the bodyfluid obtained to an A $\beta$ -affinity chromatography, and recovering the purified anti-A $\beta$  antibody. Preferably the IgG-containing bodyfluid is a fluid selected from the group consisting of cerebrospinal fluid, plasma and urine, all of them obtained from one or more human beings (pooled samples).

[0024] Furthermore, it is preferred that the A $\beta$ -affinity chromatography is carried out by an A $\beta$ -affinity column, obtained by conjugating A $\beta_{1-40}$  onto Sepharose 4B, elution with elute buffer at pH 1.5 to 2.5 at 4°C using an FPLC system.

[0025] The present invention also relates to the use of the above anti-A $\beta$  antibody and/or the use of an IgG containing, preferably IgG enriched fluid for diagnosing and/or treating amyloid associated diseases, especially Alzheimer's disease. Preferably the use is for treatment of amyloid associated diseases, especially Alzheimer's disease.

[0026] According to another embodiment there is provided a pharmaceutical composition comprising the anti-A $\beta$  antibody of the present invention. A pharmaceutical composition of the invention comprises the anti-A $\beta$  antibody and is preferably for parenteral administration, e.g. by i.v., i.m. or i.c. injection. It may comprise conventional carriers. A preferred dosage for administration is in the range of 0.001 to 3 g/kg body weight per day, a more preferred dosage for administration being in the range of 0.01 to 0.4 g/kg body weight per day.

[0027] The experimental work forming the basis of the present invention was carried out using the following materials and methods:

[0028] **A $\beta$  antibody ELISA:** 1 mg A $\beta_{(1-40)}$  is dissolved in 2ml H<sub>2</sub>O. Then add up to 200 ml coating buffer (1.7mM NaH<sub>2</sub>PO<sub>4</sub>·H<sub>2</sub>O; 98 mM Na<sub>2</sub>HPO<sub>4</sub>·7H<sub>2</sub>O, 0.05% sodium azide; pH 7.4). Add 100  $\mu$ l/well of coating buffer overnight at 4°C. Remove coating buffer and block plate with blocking buffer for 80 min. (blocking

buffer 1: 0.25% casein in PBS, 0.05% sodium azide, pH=7.4). Wash plate 3 times with washing buffer (1xPBS/0.05% Tween-20). Load samples overnight at 4°C. Remove samples and wash plate 3 times. Add monoclonal anti-human biotinylated IgG in blocking buffer 1 for 1 h. Wash 3 times with washing buffer. Load antibody against biotin conjugated with horse radish peroxidase for 1h. Wash four times and add TMB for 10 min, then add H<sub>2</sub>SO<sub>4</sub> (1N) to stop reaction and read at a plate reader at 450 nm.

[0029]  **$\beta$ -Amyloid-ELISA:** For the measurement of A $\beta$  a commercially available kit for A $\beta$ <sub>1-42</sub>, A $\beta$ <sub>1-40</sub> and A $\beta$ <sub>1-5</sub> was used.

[0030] **Cerebrospinal fluid (CSF) and plasma:** lumbar CSF and plasma were collected following standard clinical procedures after informed consent of the patients.

[0031] **Criteria for the diagnosis of Alzheimer's disease:** All normal controls had no significant decline or impairment in cognition on clinical examination. They had no history or evidence of neurological disease with potential to affect cognition and no deficits in their ability to adequately perform activities of daily living (ADLs). All AD patients had a clinical examination, including neuropsychological testing, to document deficits in cognition and ADLs, laboratory studies and a neurological examination to exclude reversible causes of dementia. All patients met ICD-10 criteria for dementia as well as NINCDS-ADRDA criteria for probable or possible AD.

#### DESCRIPTION OF THE DRAWINGS:

##### Figure 1:

[0032] A $\beta$ -antibody has been identified in the CSF from Alzheimer's disease patients (AD) and control individuals. Levels of A $\beta$  antibodies in CSF from AD patients were reduced by 30% when compared to age-matched control subjects ( $p<0.05$ , one way ANOVA)

##### Figure 2:

[0033] 1 A $\beta$  antibody unit = 10 antibody titres. 1 ml of CSF was incubated with 1, 10, or 100  $\mu$ l of protein A conjugated with agarose bead (Sigma P-7786) overnight at 4 °C (Pa). After removing protein A, 290  $\mu$ l of CSF were used to determine the titre of antibody. 1 ml of CSF was also incubated with A $\beta$ <sub>1-40</sub>, A $\beta$ <sub>1-42</sub>, A $\beta$ <sub>25-35</sub> (1 mg A $\beta$  was dissolved in 0.9 ml of deionised H<sub>2</sub>O). 0.8-20  $\mu$ L of A $\beta$  was used for overnight incubation (at 4 °C) with CSF. 290 ml of CSF was then used for determination.

PE 1-100  $\mu$ L: Protein A precipitates (1, 10, or 100  $\mu$ l) from CSF sample was incubated with 100  $\mu$ l of PBS (pH 2.5). The recovered solution was used for titre determination. The antibody titre is defined as the dilution of antibody that gives a half-maximal binding to antigen. (Pa: Protein A; A $\beta$ <sub>1-40</sub>:  $\beta$ -Amyloid 1-40; A $\beta$ <sub>1-42</sub>:  $\beta$ -Amy-

loid 1-42; A $\beta$ <sub>25-35</sub>:  $\beta$ -Amyloid 25-35; PE: elute from protein A precipitates)

##### Figure 3:

[0034] Same condition as in Figure 2. Only A $\beta$ <sub>1-40</sub>=2  $\mu$ l. A $\beta$ <sub>40-1</sub>=2  $\mu$ l. Neuropeptide F and neuropeptideY (2  $\mu$ l), Amylin (2  $\mu$ l).

##### Figure 4:

[0035] Purification of anti-A $\beta$  antibodies by using A $\beta$  affinity column.

After 250 g immunoglobulin (IgG) pass through the A $\beta$  affinity column, 10 ml of elute buffer (pH2.5) was used to elute A $\beta$  antibody. Then another 10 ml of elute buffer (pH1.5) was used to elute the remainder of antibodies. After ELISA detection, significant amount of A $\beta$  antibody was detected in pH 2.5 elute buffer. IgG = immunoglobulin 100  $\mu$ l. PH2.5, 1.0: elute antibodies by using pH2.5 and then pH1.5 buffers from affinity column: 100  $\mu$ l. PT: IgG pass through A $\beta$  affinity column, equal to 100  $\mu$ l of IgG. Most anti-A $\beta$  antibody elute from column by pH 2.5. Column: 3 mg of A $\beta$ <sub>1-40</sub> was conjugated into Sepharose 4B (Pharmacia, 5 ml). Purification by using Pharmacia FPLC system at 4 degree. 1 A $\beta$  antibody unit = 10 antibody titres. (Elute buffer: 50 mM glycine, 150 mM NaCl, pH 2.5).

##### Figure 5:

[0036] Concentration of  $\beta$ -Amyloid in the CSF before treatment with immunoglobulins and 7-12 days and 4 weeks after treatment, respectively. Measurements were done as described in Example 2.

[0037] The following examples are given for illustration purposes only and are not intended to limit the scope of the invention.

[0038] Example 1: Treatment of AD patients by infusion of human IgG Immunoglobulins or anti-A $\beta$  antibodies from human IgG.

[0039] As an example as to the therapy regimen 5 - 30 g (1 - 5 days) of IgG immunoglobulins (commercially available) or a corresponding amount of purified anti-A $\beta$  antibody are administered parenterally to the patient by the i.v. route. Levels of  $\beta$ -Amyloid, tau-protein as well as A $\beta$ -antibody are measured in the serum and CSF before and following the respective dose of IgG immunoglobulins for therapy control. The goal is to decrease  $\beta$ -amyloid concentration in the CSF and by that decrease the plaque burden in Alzheimer's disease and alleviate the neuropsychiatric and neuropsychological defects in Alzheimer's disease. This treatment introduces a new therapeutic approach to Alzheimer's disease.

**Example 2: Effect of i.v. immunoglobulins on the Concentration of  $\beta$ -Amyloid in the CSF**

[0039] In this example the effect of the application of i.v.-immunoglobulins (Octagam®, Polyglobulin®) on the concentration of  $\beta$ -Amyloid in the CSF is investigated.

[0040] Four patients suffering from different neurological disorders (Guillain-Barre-Syndrome; chronic inflammatory demyelinating neuropathy, CIDP) were included in this study. Lumbar CSF was withdrawn before starting treatment with i.v. immunoglobulins. After 7 to 12 days and 4 weeks an additional lumbar puncture was performed. The withdrawal of CSF was performed during regular investigations. Patients were treated with i.v. immunoglobulins for 3 - 5 days with 0.4 g/kg per day before withdrawal of CSF. The concentration of  $\beta$ -Amyloid was measured in the CSF before treatment and 7 - 12 days and 4 weeks after application of i.v. immunoglobulins. The results are shown in Fig. 5.

[0041] From the figure it can be seen that the amount of  $\beta$ -Amyloid was reduced from 1835 ng/l before treatment to 1622 ng/l (7-12 d after treatment) and 1376 ng/l (4 weeks after treatment). These results show that i.v. administration of immunoglobulins has an effect on the concentration of  $\beta$ -Amyloid in the CSF. Immunoglobulins also reduce  $\beta$ -Amyloid in the brain of patients with Alzheimer's disease.

tained by conjugating  $A\beta_{1-40}$  onto Sepharose 4B, elution with elute buffer at pH 1.5 to 2.5 at 4°C using an FPLC system.

- 5 6. Use of an anti- $A\beta$ -amyloid antibody according to claims 1 or 2 for treating amyloid associated diseases, especially Alzheimer's disease and primary and secondary amyloidoses.
- 10 7. Use of an anti- $A\beta$ -amyloid antibody according to claims 1 or 2 for diagnosis of amyloid associated diseases, especially Alzheimer's disease and primary and secondary amyloidoses.
- 15 8. Use of an IgG containing, preferably IgG enriched fluid for treatment of amyloid associated diseases, especially Alzheimer's disease.
- 20 9. Pharmaceutical composition comprising an anti- $A\beta$ -amyloid antibody according to claims 1 or 2.

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**Claims**

- 1. A human anti- $A\beta$ -amyloid antibody obtained by purification from a human IgG-containing bodyfluid by  $A\beta$ -affinity chromatography. 35
- 2. Human anti- $A\beta$ -amyloid antibody of claim 1, characterised in that it belongs to the class of immunoglobulines G (IgG) and does not recognise  $A\beta_{40-1}$ , neuropeptide F, neuropeptide Y, and Amylin, and specifically recognises one or more of  $A\beta_{1-40}$ ,  $A\beta_{1-42}$  and  $A\beta_{25-35}$ . 40
- 3. A method of purification of an anti- $A\beta$ -amyloid antibody comprising the steps of 45
  - (i) obtaining a human IgG-containing bodyfluid,
  - (ii) subjecting the bodyfluid obtained to an  $A\beta$ -affinity chromatography, and
  - (iii) recovering the purified anti- $A\beta$  antibody. 50
- 4. The method of claim 3 wherein the IgG-containing bodyfluid is a cerebrospinal fluid, plasma or urine obtained from one or more human beings (pooled samples). 55
- 5. The method of claim 3, wherein  $A\beta$ -affinity chromatography is carried out by an  $A\beta$ -affinity column, ob-

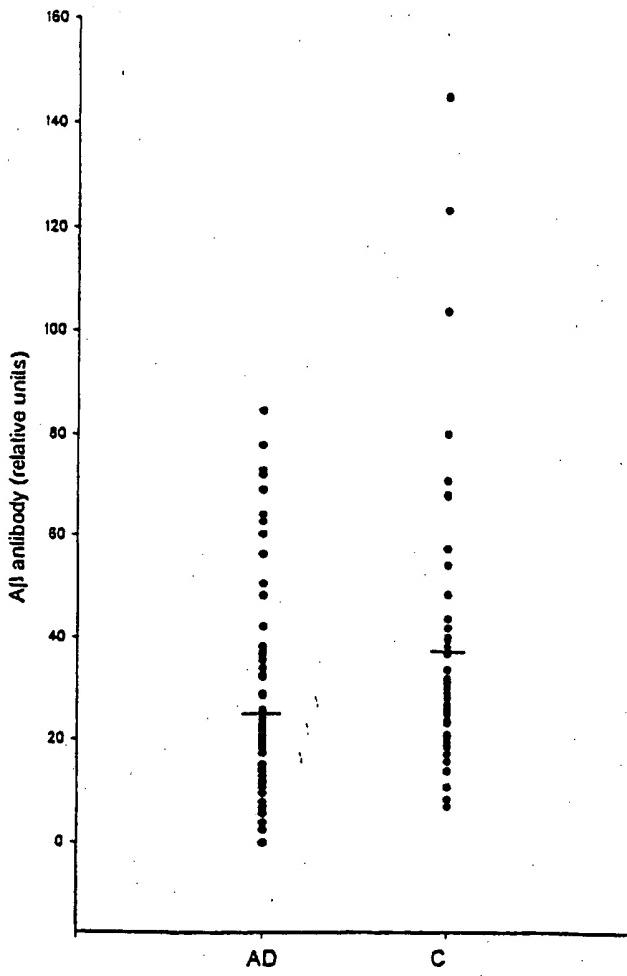


Figure 1

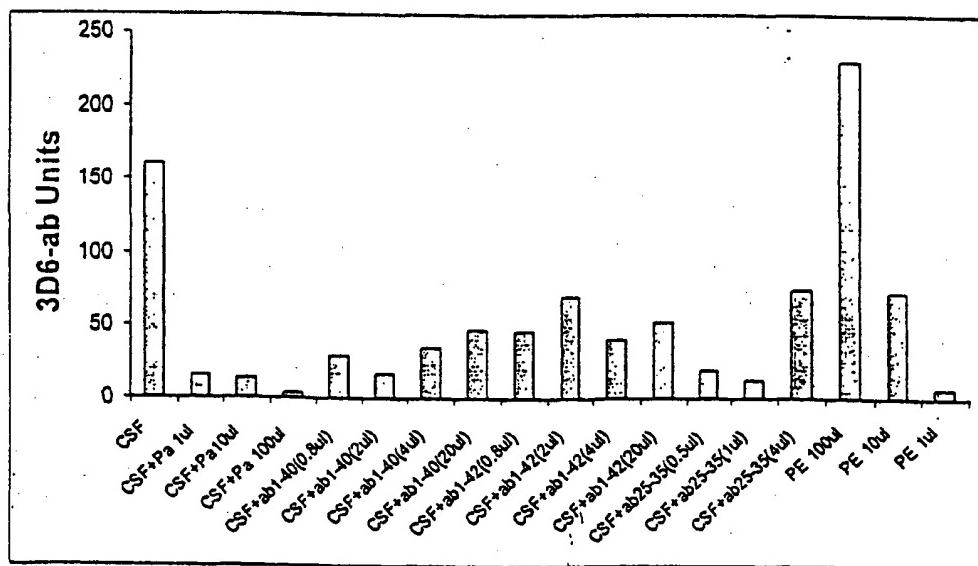


Figure 2

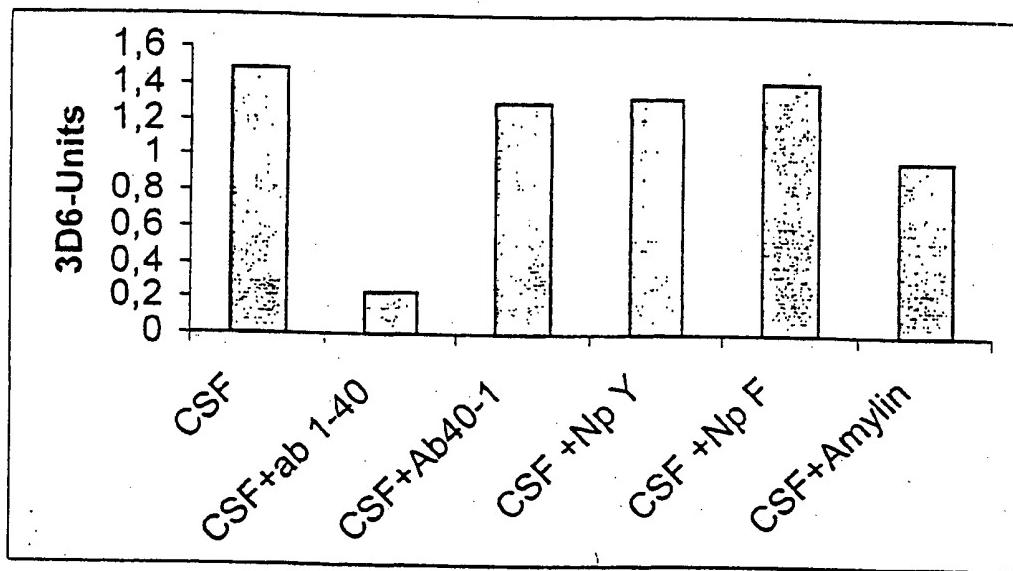


Figure3

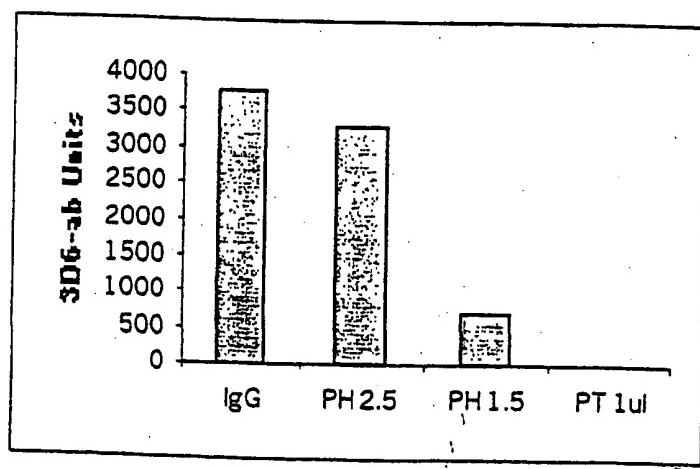


Figure 4

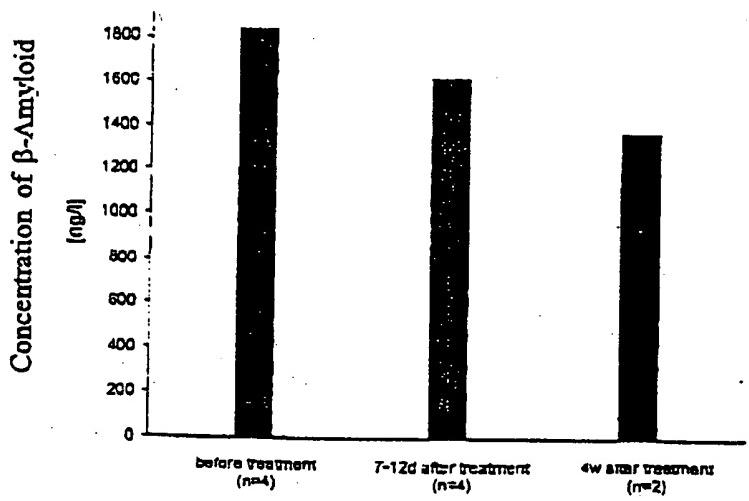


Figure 5:



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## EUROPEAN SEARCH REPORT

Application Number  
EP 01 11 4825

DOCUMENTS CONSIDERED TO BE RELEVANT									
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim	CLASSIFICATION OF THE APPLICATION (Int.Cl.)						
A	GASKIN F ET AL: "Human antibodies reactive with beta- amyloid protein in Alzheimer 's disease." JOURNAL OF EXPERIMENTAL MEDICINE, (1993 APR 1) 177 (4) 1181-6. , XP001030627 * the whole document *	1-9	C07K16/18 A61K39/395 C07K1/22 G01N33/68 A61P25/28						
A	XU S ET AL: "Increased incidence of anti-beta- amyloid autoantibodies secreted by Epstein-Barr virus transformed B cell lines from patients with Alzheimer 's disease." MECHANISMS OF AGEING AND DEVELOPMENT, (1997 MAR) 94 (1-3) 213-22. , XP001030610 * the whole document *	1-9							
A	SEUBERT P ET AL: "ISOLATION AND QUANTIFICATION OF SOLUBLE ALZHEIMER'S BETA-PEPTIDE FROM BIOLOGICAL FLUIDS" NATURE, MACMILLAN JOURNALS LTD. LONDON, GB, vol. 359, no. 6393, 24 September 1992 (1992-09-24), pages 325-327, XP000616173 ISSN: 0028-0836 * page 326, right-hand column - page 327, right-hand column *	1-9							
A	WO 83 00048 A (THE BETH ISRAEL HOSPITAL ASSOCIATION) 6 January 1983 (1983-01-06) * claims 1-10 *	1-9 -/-	C07K A61K						
<p>The present search report has been drawn up for all claims</p> <table border="1" style="width: 100%; border-collapse: collapse;"> <tr> <td style="width: 33%;">Place of search</td> <td style="width: 33%;">Date of completion of the search</td> <td style="width: 33%;">Examiner</td> </tr> <tr> <td>THE HAGUE</td> <td>13 November 2001</td> <td>Le Flao, K</td> </tr> </table>				Place of search	Date of completion of the search	Examiner	THE HAGUE	13 November 2001	Le Flao, K
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DOCUMENTS CONSIDERED TO BE RELEVANT			CLASSIFICATION OF THE APPLICATION (Int.Cl.)															
Category	Citation of document with indication, where appropriate, of relevant passages	Relevant to claim																
P, X	FRENKEL, DAN ET AL: "Immunization against Alzheimer's beta-amyloid plaques via EFRH phage administration" PROC. NATL. ACAD. SCI. U. S. A. (2000), 97(21), 11455-11459 , XP002180061 * the whole document *	1-9																
P, X	SOLOMON B. ET AL: "Vaccination for the prevention and treatment of Alzheimer's disease." DRUGS OF TODAY, (2000) 36/9 (655-663). , XP001030591 * the whole document *	1-9																
			TECHNICAL FIELDS SEARCHED (Int.Cl.)															
<p>The present search report has been drawn up for all claims</p> <table border="1"> <tr> <td>Place of search</td> <td>Date of completion of the search</td> <td>Examiner</td> </tr> <tr> <td>THE HAGUE</td> <td>13 November 2001</td> <td>Le Flao, K</td> </tr> <tr> <td colspan="3">CATEGORY OF CITED DOCUMENTS</td> </tr> <tr> <td colspan="3">           X : particularly relevant if taken alone            Y : particularly relevant if combined with another document of the same category            A : technological background            O : non-written disclosure            P : intermediate document         </td> </tr> <tr> <td colspan="3">           T : theory or principle underlying the invention            E : earlier patent document, but published on, or after the filing date            D : document cited in the application            L : document cited for other reasons            &amp; : member of the same patent family, corresponding document         </td> </tr> </table>				Place of search	Date of completion of the search	Examiner	THE HAGUE	13 November 2001	Le Flao, K	CATEGORY OF CITED DOCUMENTS			X : particularly relevant if taken alone Y : particularly relevant if combined with another document of the same category A : technological background O : non-written disclosure P : intermediate document			T : theory or principle underlying the invention E : earlier patent document, but published on, or after the filing date D : document cited in the application L : document cited for other reasons & : member of the same patent family, corresponding document		
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Patent document cited in search report	Publication date	Patent family member(s)	Publication date
WO 8300048 A	06-01-1983	US 4722896 A	02-02-1988
		CA 1193195 A1	10-09-1985
		EP 0081583 A1	22-06-1983
		WO 8300048 A1	06-01-1983

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